CLEAN VERSION OF REPLACEMENT PARAGRAPHS
IN THE SPECIFICATION PURSUANT TO 37 C.F.R. §1.121(b)

IN THE SPECIFICATION

On page 8, please delete the paragraphs beginning on line 23 and ending on line 31, and replace with the following paragraphs:

A1

Figure 15 shows the nucleic acid sequence of the promoter region P6 of gene 6 (SEQ ID NO: 25), with an inverted repeat indicated by highlight (SEQ ID NO: 26). The BLAST result of the sequence blasted against its reverse complementary sequence is also shown.

Figure 16 shows the nucleic acid sequence of the promoter region P14 of gene 14 (SEQ ID NO: 27), with an inverted repeat indicated by highlight (SEQ ID NO: 28). The BLAST result of the sequence blasted against its reverse complementary sequence is also shown.

Figure 17 shows the nucleic acid sequence of the promoter region P16 of gene 16 (SEQ ID NO: 29), with an inverted repeat indicated by highlight (SEQ ID NO: 30). The BLAST result of the sequence blasted against its reverse complementary sequence is also shown.

On page 68, please delete the paragraph beginning on line 21 and ending on page 69, line 17, and replace with the following paragraph:

A2

The plasmids of 2715 selected cDNA clones were collected from data set I. The inserts of the cDNAs were amplified by PCR in a 96-well format using primer pairs specific for the vector ends (for inserts in pBluescript SK-: T7, 5′-GTAATACGACTCACTATAGGGC (SEQ ID NO: 55), and 5′ extended M13 reverse, 5′-ACAGGAAACAGCTATGACCATG (SEQ ID NO: 56); for inserts in pZipLox1: M13 forward, 5′-CCCAGTCACGACGTTGTAAAACG (SEQ ID NO: 57) and M13 reverse, 5′-AGCGGATAACAATTTCACACAGG (SEQ ID NO: 58). PCR reactions of 100 µL volume contained 0.4 µM of each primer, 0.2 µM of each desoxynucleotide, 10 mM Tris, 50 mM KCl, 3.0 mM $MgCl_2$, 3 U *Taq* DNA polymerase (Promega, Madison) and ~10 ng plasmid template. The reactions were run on a Perkin Elmer 9700 Thermoblock using an amplification program of 3 min denaturation at 94 °C, 5 precycles of 30 s at 94 °C, 30 s at 64 °C, 2 min at

72 °C, followed by 30 cycles of 30 s at 94 °C, 30 s at 60 °C, 2 min at 72 °C and terminated by 7 min extension at 72 °C. The PCR products were precipitated by adding 200 µL ethanol (95%) and 10 µL sodium acetate (3M, pH 5.2) and centrifugation at 3200 g and 4 °C for 60 min. After washing with 80% ethanol, the DNA was resuspended in 20 µL 3x SSC. The yield and purity of the PCR products was analyzed by agarose gel electrophoresis. PCR samples showing by agarose gel analysis concentrations less than 0.2 µg/µL and/or double bands were repeated. If possible, alternative clones from the cDNA clone collection were used to repeat the PCR experiments. To reduce the cross-contamination risk in the 96-well format, failed PCRs were not removed from the sample set, and as a result the number of PCR samples for printing increased by approximately 20%.

A2 cancel

On page 74, please delete the paragraph beginning on line 19 and ending on line 28, and replace with the following paragraph:

A3

Control vectors contained a GUS expression vector with either a napin or phaseolin promoter. For example, the promoter region of the napin (napA) gene in Brassica napus was amplified by using a forward primer CG aagctt TCTTCATCGGTGATT (SEQ ID NO: 59) and reverse primer GGTCG gaattc GTGTATGTTTT (SEQ ID NO: 60). The PCR product was digested by Hind III and EcoR I, then inserted into SK+ vector and confirmed by sequencing. The napin promoter was cut by Hind III and BamH I and inserted into a GUS expression vector such that GUS is under control of the napin promoter region. In a similar fashion, a GUS expression vector under control of a phaseolin promoter region was constructed; the phaseolin promoter region is described in patent US 5,504,200.

Please insert the attached Sequence Listing as new pages --80-107--.

IN THE FIGURES:

Please replace Figures 19a, 19b, and 19c with the attached Figures 19a, 19b, and 19c Substitute Sheets.

IN THE CLAIMS:

Please renumber the Claims pages from pages "80-84" to --108-112--.